## SPECIAL REPORT

## The role of the $I_{sK}$ protein in the specific pharmacological properties of the $I_{Ks}$ channel complex

<sup>1</sup>A.E. Busch, G.L. Busch, E. Ford, H. Suessbrich, †H.-J. Lang, \*R. Greger, \*K. Kunzelmann, #B. Attali & W. Stühmer

Max-Planck-Institut für experimentelle Medizin, Hermann-Rein-Str. 3, D-37075 Göttingen, †Hoechst AG, D-65926 Frankfurt/Main, \*Physiological Institute, Hermann-Herder-Str. 7, D-79140 Freiburg, Germany and #Department of Neurobiology, The Weizmann Institute of Science, Rehovot, 76100, Israel

 $I_{\rm Ks}$  channels are composed of  $I_{\rm sK}$  and KvLQT1 subunits and underly the slowly activating, voltage-dependent  $I_{\rm Ks}$  conductance in heart. Although it appears clear that the  $I_{\rm sK}$  protein affects both the biophysical properties and regulation of  $I_{\rm Ks}$  channels, its role in channel pharmacology is unclear. In the present study we demonstrate that KvLQT1 homopolymeric K<sup>+</sup> channels are inhibited by the  $I_{\rm Ks}$  blockers 293B, azimilide and 17- $\beta$ -oestradiol. However,  $I_{\rm Ks}$  channels induced by the coexpression of  $I_{\rm sK}$  and KvLQT1 subunits have a 6–100 fold higher affinity for these blockers. Moreover, the  $I_{\rm Ks}$  activators mefenamic acid and DIDS had little effect on KvLQT1 homopolymeric channels, although they dramatically enhanced steady-state currents through heteropolymeric  $I_{\rm Ks}$  channels by arresting them in an open state. In summary, the  $I_{\rm sK}$  protein modulates the effects of both blockers and activators of  $I_{\rm Ks}$  channels. This finding is important for the action and specificity of these drugs as  $I_{\rm sK}$  protein expression in heart and other tissues is regulated during development and by hormones.

Keywords: Heart; arrhythmia; antiarrhythmics; K channel; chromanol

Introduction The  $I_{sK}$  (also called minK) protein induces slowly activating, voltage-dependent K<sup>+</sup> channels in *Xenopus* oocytes, previously called  $I_{\min K}$ ,  $I_{sK}$  or  $I_{Ks}$  channels (Takumi et al., 1988). Iks channels in Xenopus oocytes exhibit almost identical biophysical, pharmacological and regulatory properties as described for the  $K^+$  conductance  $I_{Ks}$  in cardiac myocytes (reviewed by Busch & Suessbrich, 1997). Recent studies showed that the  $I_{sK}$  protein heteropolymerizes with a 'classical'  $K^+$  channel subunit (KvLQT1) to form functional I<sub>Ks</sub> channels (Sanguinetti et al., 1996; Barhanin et al., 1996). I<sub>Ks</sub> channels represent a potentially important target for antiarrhythmic drugs. It is important to elucidate the individual roles of  $I_{sK}$  and KvLQT1 subunits for drug binding within the  $I_{Ks}$ channel complex. In the present study we therefore analysed the effects of distinct  $I_{Ks}$  channel blockers and activators on mouse KvLQT1 subunits expressed either alone or together with the human  $I_{sK}$  protein to form heteromultimeric  $I_{Ks}$ channels.

**Methods** Handling of *Xenopus* oocytes, synthetis of cRNA, voltage-clamp experiments and the analysis thereof have been described in detail (Busch *et al.*, 1994a). Azimilide was a gift from Procter & Gamble Pharmaceuticals. 17- $\beta$ -oestradiol, DIDS (4,4'-diisothiocynostilbene-2,2'-disulphonic acid) and mefenamic acid were purchased from Sigma. Concentration-blockade releationships were calculated with the Hill equation. Student's *t* test was used to test for statistical significance, which was assumed if *P*<0.05.

**Results** Expression of mouse KvLQT1 in *Xenopus* oocytes alone or together with the  $I_{\rm sK}$  protein induced voltage-dependent K<sup>+</sup> channels (Figure 1a) with similar characteristics as previously described (Barhanin *et al.*, 1996). The current amplitude after 2 days of  $I_{\rm sK}/{\rm KvLQT1}$  coexpression (i.e.  $I_{\rm Ks}$ 

channels) was approximately 4 fold larger than the current induced by expression of the  $I_{\rm sK}$  protein alone (which forms  $I_{\rm Ks}$  channels with endogenous KvLQT1 subunits in *Xenopus* oocytes; Sanguinetti *et al.*, 1996). However, the possibility that heteropolymerization of endogenous and exogenous KvLQT1 subunits may occur for a small portion of the KvLQT1 and  $I_{\rm Ks}$  channel populations cannot be excluded.

Oocytes expressing KvLQT1 channels or  $I_{Ks}$  channels were analysed for their sensitivity to the blockers 293B, azimilide and 17-β-oestradiol (Busch et al., 1994b; Busch et al., 1996; Waldegger et al., 1996). 293B, azimilide and  $17-\beta$ -oestradiol blocked KvLQT1 with estimated IC<sub>50</sub> values of  $40.9 \pm 0.9 \mu M$ (n=5), 77.4 ± 6.9  $\mu$ M (n=5) and > 50  $\mu$ M (n=5), respectively. Coexpression of KvLQT1 and  $I_{sK}$  subunits induced  $I_{Ks}$  channels which were blocked by 293B, azimilide and 17-β-oestradiol with IC<sub>50</sub> values of  $6.7 \pm 0.5 \mu M$  (n = 5),  $5.6 \pm 0.7 \mu M$  and  $2.2 \pm 1.0 \ \mu \text{M}$  (Figure 1b; n = 5), respectively. Therefore, the coexpression of  $I_{sK}/KvLQT1$  subunits induced  $I_{Ks}$  channels with a much higher blocker affinity compared to homopolymeric KvLQT1 channels. Moreover, the  $I_{Ks}$  channels induced by the overexpression of both  $I_{sK}$  and exogenous KvLQT1 subunits exerted the same sensitivity to the applied blockers as previously described for  $I_{Ks}$  channels induced by the expression of the  $I_{sK}$  protein alone. This suggests that exogenous and endogenous KvLQT1 subunits possess a similar affinity for these compounds, which is no surprise considering the high homology of the two proteins.

The Cl $^-$  channel blockers mefenamic acid and DIDS are known to have positive modulatory effects on  $I_{\rm Ks}$  channels induced by expression of the human  $I_{\rm sK}$  protein alone (Busch *et al.*, 1994a). As shown in Figure 2a and b, mefenamic acid (0.1 mM) had little effect on KvLQT1 channels (n=5). In contrast, the coexpression of  $I_{\rm sK}$  and KvLQT1 subunits induced  $I_{\rm Ks}$  channels with high sensitivity to mefenamic acid. As shown in Figure 2a and b, superfusion with mefenamic acid increased steady state currents at -10 mV about 5 fold and arrested  $I_{\rm Ks}$  channels in an open state, i.e. at the holding potential of -70 mV no significant  $I_{\rm Ks}$  channel deactivation was observed (n=5). This effect was rapidly reversed upon wash-out. Qualitatively similar results on  $I_{\rm Ks}$  deactivation and steady state currents were observed with 0.1 mM DIDS (Figure 2b; n=5).

<sup>&</sup>lt;sup>1</sup> Author for correspondence at present address: Physiological Institute, Gmelinstr.5, D-72076 Tübingen, Germany.

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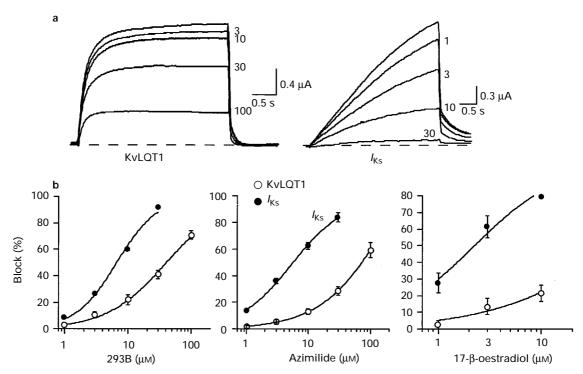
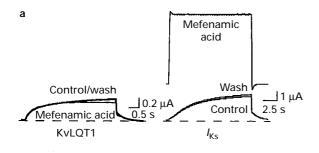
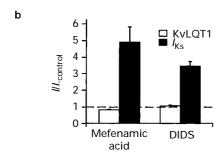


Figure 1 Effects of  $I_{Ks}$  blockers on KvLQT1 and  $I_{Ks}$  channels in *Xenopus* oocytes. (a) Left: KvLQT1 expression induced fast activating potassium channels (depolarizations to  $-20\,\text{mV}$  from a holding potential of  $-70\,\text{mV}$ ) Right:  $I_{Ks}$  channels were slower at activating than KvLQT1 channels and were more sensitive to 293B.  $I_{Ks}$  channels were activated with depolarizations from  $-70\,\text{mV}$  to  $-10\,\text{mV}$ . (b) Concentration-dependence of inhibition by 239B, azimilide and 17-β-oestradiol of KvLQT1 and  $I_{Ks}$ .





**Figure 2** (a) Effects of mefenamic acid (0.1 mm) on KvLQT1 (left) and  $I_{\rm Ks}$  channels (right).  $I_{\rm Ks}$  channels were repetitively activated with depolarizations from  $-70\,{\rm mV}$  to  $-10\,{\rm mV}$ . (b) Effects of mefenamic acid and DIDS (0.1 mm) on steady state currents at  $-10\,{\rm mV}$  of KvLQT1 and  $I_{\rm Ks}$ . The currents were normalized against control currents. The dashed line represents the relative control current.

**Discussion** The present data demonstrate that the  $I_{sK}$  protein plays a crucial role not only for the biophysical properties and regulation of  $I_{Ks}$  channels but also for their pharmacology. Upon overexpression it enhances the sensitivity to blockers and even more dramatically to  $I_{Ks}$  activators. This suggests that the  $I_{sK}$  protein contributes to the binding of these compounds. The binding region for these compounds may also represent a crucial interaction interface between  $I_{sK}$  and KvLQT1 subunits. Both IsK and KvLQT1 proteins are expressed in heart and numerous other epithelial tissues (reviewed by Busch & Suessbrich, 1997). Interestingly, a study by Folander et al. (1990) suggested that the sex hormone oestrogen upregulates  $I_{sK}$  protein expression in uterus, whereas Drici et al. (1996) found a downregulation in the heart after oestrogen treatment. A differential expression of the  $I_{sK}$  and KvLQT1 subunits may therefore also account for the tissuespecific action of drugs such as those described in the present study.

A.E.B. and K.K. are Heisenberg fellows, G.L.B. is a Habilitation fellow of the Deutsche Forschungsgemeinschaft (DFG), respectively. The work was supported by the DFG (Bu 704/3-2) and the German Israel Foundation. The authors appreciate the help of Drs M. Stocker and P. Pedarzani. We thank Dr M. Lazdunski for providing the KvLQT1 clone.

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(Received June 9, 1997 Accepted July 15, 1997)